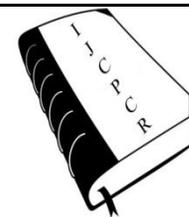




International Journal of Current Pharmaceutical & Clinical Research



www.ijcpcr.com

NATURAL REMEDIES: THE FUTURISTIC THERAPY FOR ANTIBIOTIC RESISTANT SUPERBUGS

Govind shukla*, Sangeeta kumari ¹, Dr.Vinod Yadav ², Dr.S.B Dixit ³, Dr.Manjula Dixit ⁴,
KelashKumar ⁵, Marlon O. Perez ⁶, Mercedita H. Perez ⁷, Ratish ShyamDixit ⁸,
V. Chandramauli ⁹

¹Research Scholar, JNTU, Hyderabad, India.

²Chief Medical Officer, Jhansi, Uttar Pradesh, India.

³Professor & Head, Department of Community Medicine, Manipal college of Medical sciences Pokhra NEPAL

⁴Professor, Department of Pathology, Manipal College of Medical Sciences, Pokhra, Nepal.

⁵CPO QA, Specialist Novartis Pharma Limited, Karachi, Pakistan

⁶Graduate School Student, West Visayas State University, Iloilo City, Philippines.

⁷RT II Ultrasound Section Head, Diagnostic Imaging Department & Radiological Sciences, Corazon Locsin Montelibano Memorial Regional Hospital, Lacson St., Bacolod City, Philippines.

⁸Manager Marketing (Head), BIOCON Ltd, Bangalore, India.

⁹Consultant Physician & Nutritionist, Praja Vaidyashala, P&T Colony, Dilsukh Nagar, Hyderabad, India.

ABSTRACT

Imagine being sick in the hospital with a bacterial infection and doctors can't stop it from spreading. This so-called "superbug" scenario is not science fiction. It's an urgent, worldwide worry that is prompting swift action. Antibiotic-resistant illnesses currently kill an estimated 700,000 people a year globally. By 2050, these illnesses are expected to kill 10 million people. Based on recent research, it could be even worse—and coming even sooner. Ancient remedies, including essential oils and their components, have been explored as a source of new antimicrobials. Many are known to possess significant antimicrobial activity against a wide range of microorganisms. Additionally, combination of existing drugs with essential oils and/or components may provide an alternative approach to combat emerging drug resistance. Since antibiotic resistance is currently outpacing research and development to find new drugs, humanity is facing a return to the 'pre-antibiotic era'. Perhaps the remedies of the past combined with scientific study may provide the antibiotics of tomorrow. The present paper emphasized the role of Natural Remedies for Antibiotic Resistant superbugs.

Key words: Natural Remedies, Antibiotic Resistant superbugs, Futuristic therapy.

INTRODUCTION

Sir Alexander Fleming discovered the antibacterial power of the mold *Penicillium notatum* in 1928. Even though it was a natural healing agent effective in destroying *Staphylococcus aureus* and other noxious bacteria, Because of the overuse of antibiotics, super-bugs have developed that are resistant to all but the most powerful drugs, whose side effects are often dangerous.

Antibiotic resistance is emerging at an alarming rate, outpacing current research and development efforts to combat this trend. As a result, many infectious diseases have become difficult to treat; in some cases, no treatment options exist. The search for new antibiotics must accelerate to avoid returning to the 'pre-antibiotic' era. Ancient remedies, including essential oils and their

Corresponding Author :- **Govind shukla** Email:- govindbbd@gmail.com

components, have been explored on a limited basis as a source of new antimicrobials. Many are known to possess significant antimicrobial activity against a wide range of microorganisms.

How antibiotic-resistant bacteria can spread

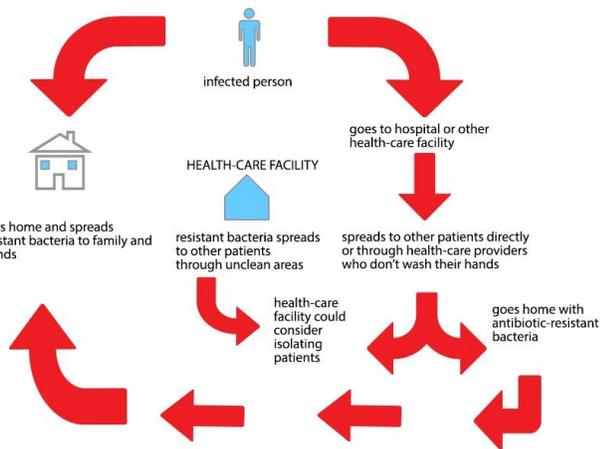


Fig 1.

Elucidation of the mechanism of action of these compounds may lead to identification new antibiotic targets. Such targets, once identified, may represent biosynthetic or regulatory pathways not currently inhibited by available drugs.

Novel drugs and targets are vital for continued control of infectious diseases worldwide. In nature's infinite wisdom, several other highly effective substances exist with antibacterial, antifungal and antiviral properties, all able to protect the human body safely and with deep healing powers [1,2].

Natural Remedies and superbugs

For thousands of years, ancient remedies have been used to relieve a wide variety of human maladies including bronchitis, pneumonia, pharyngitis, diarrhea, periodontal disease, wounds, and numerous other illnesses. Many traditions surrounding the use of these oils are buried in antiquity, passed down orally from master to student until the origin of specific treatments were lost to the ages. In antiquity, medicinal oils were derived from aromatic plants and resins by extraction into other fatty oils such as olive oil and used as a mixture. Ancient remedies, including essential oils and their components, have been explored on a limited basis as a source of new antimicrobials. Many are known to possess significant antimicrobial activity against a wide range of microorganisms.

The earliest recorded use of aromatic oils dates back to 4,500 B.C. in Egypt [3]. The ancient Egyptians recognized that oils could be used in treating illness, including infection and inflammation. So valuable were these oils, that King Tutankhamun was entombed with roughly 350 liters of aromatic oil including cedarwood, frankincense, and myrrh [3]. Myrrh is one of the earliest and well known of the aromatic oils. The ancient Hebrews referred to myrrh as 'holy oil' which was more valuable than gold. The ancient Egyptians referred to myrrh as 'the tears of Horus'. Myrrh is derived from the resin of a woody shrub of the genus, *Commiphora*, which grows in hot, arid climates. In ancient Sumer, myrrh was used for treating parasitic infections and periodontal disease. The Greek physician, Dioscorides, used myrrh for bronchial and other infections including the skin [4]. Myrrh was often combined with frankincense, aromatic oil used in antiquity to treat infectious diseases and inflammation. Like myrrh, frankincense is member of a resinous family of plants (*Burseraceae*) commonly found in arid regions of the Middle East and north-east Africa [5,6]. The use of frankincense and myrrh is mentioned numerous times in biblical and other ancient texts [7]. These oils, alone or in combination, were used extensively for the treatment of wounds, inflammation, cystitis, rheumatic joints, skin sores, bleeding, fungal infections, burns, pharyngitis, syphilis, and leprosy [8,9].

Other cultures across the globe have long-standing, medical practices which incorporate the use of aromatic oils and other plant-based therapies, including those found in the Americas, Australia, and the Far East such as the Ayurvedic, Unani, and Chinese traditions. Among the more well-recognized remedies still in use today from North and South America and Australia are purple coneflower (*Echinacea purpurea*), cat's claw (*Uncaria tomentosa*), and eucalyptus (*Eucalyptus globulus*) [4]. Ayurvedic traditions include the use of camphor (*Cinnamomum camphora*) and cardamomum (*Elettaria cardamomum*) [4]. China's use of herbal medicine dates as far back as 3000 B.C., when the mythological and

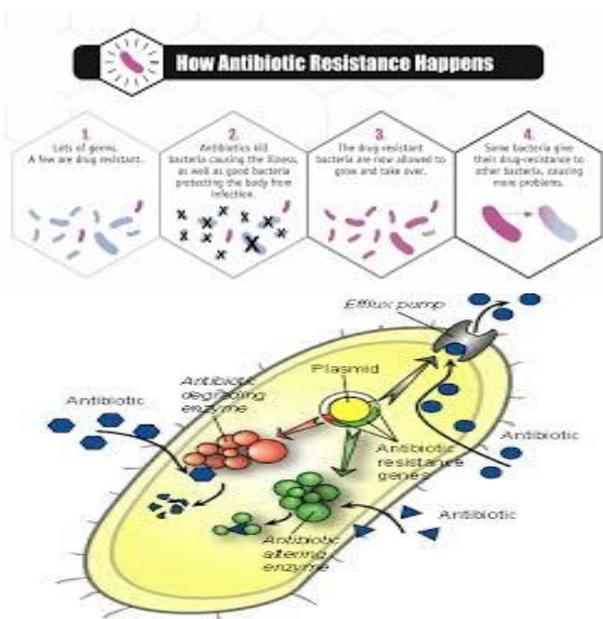


Fig 2.

legendary ruler Shen Nong Shi (or Shennong) taught humans the use of medicinal plants. His cumulative work, 'Shennong Bencao Jing', is considered one of the earliest medical collections in China [10]. By 500 A.D. the use of aromatic oils had spread throughout most of Asia Minor, and the Mediterranean, spreading along with the Roman and later the Persian Empires [11]. Commonly used aromatic oils included those derived from thyme, clove, rosemary, lavender, and cinnamon.

Today, the term 'essential oils' is used to describe the mixtures derived from aromatic medicinal plants using conventional techniques such as distillation and chromatographic separation. These oils continue to be used for the treatment of infectious disease and inflammation in traditional medicine across the globe. They are administered orally, topically, or via aromatherapy, depending on historical use and chemical composition which for many essential oils has been determined. As a result, a significant amount of toxicity data is available for not only the oils but also the individual components such that many are generally regarded as safe (GRAS) by the FDA. GRAS status has permitted the use of essential oils as flavoring agents in food and as additives to cosmetics, perfumes, and cleaning products.

The Science of natural Remedies

Essential oils are derived from a variety of natural sources including plants or components of plants such as flowers, leaves, bark, roots, berries, seeds and/or fruit. These oils are complex mixtures of chemicals, and include various alcohols, aldehydes, terpenes, ethers, ketones, phenols, and oxides. Many essential oils have limited solubility in aqueous solutions and form emulsions with non-ionic surfactants.

Previous investigators have reviewed the effect of essential oils, their components and antimicrobial activity [12-17]. However, few studies have determined the antimicrobial-specific mechanism(s) of action of various essential oils or their components [18,19].

Since essential oils are complex mixtures of compounds, it is likely the observed antimicrobial activity is due to inhibition or interaction with multiple targets in the cell [20,21]. However, many essential oils exert non-specific antimicrobial effects due to the hydrophobic properties of the mixtures and components. For instance, the hydrophobic character of many essential oils facilitates entry into cell membranes leading to alteration in architecture, leakage of cell contents, and eventually death [22-26]. In 2009, Fisher and Phillips demonstrated uptake of

Citrus sinensis and *Citrus bergamia* oils into *Enterococcus faecium* and *E. faecalis* resulting in multiple membrane-related changes: a 2- or 40- fold increase in membrane permeability, a decrease in intracellular pH, the loss of membrane potential, and a reduction in ATP concentration [25]. many essential oils contain high

concentrations of phenolic compounds including carvacrol, thymol, and eugenol. Phenols are known to disrupt cell membranes resulting in the dissolution of the proton motive force and a subsequent decrease in ATP synthesis [27-29]. Inhibition of ATP synthesis may also result from essential oil-mediated alteration of protein-protein interactions in the cell membrane or direct binding of oil components, especially cyclic hydrocarbons, to lipophilic regions of membrane-bound proteins [28,30]. Diminished ATP levels would necessarily lead to reduction in other energy-dependent cellular processes including synthesis of enzymes and toxins. For example, previous studies have demonstrated a significant decrease in the amount of diarrheal toxin detected in *Bacillus cereus* when exposed to carvacrol. The authors hypothesized that the decrease in toxin detection may be connected to the decrease in ATP production which is required not only for toxin synthesis but also export [31].

Although the spectrum of activity for most essential oils is relatively broad, as would be expected with a mechanism of action related to membrane disruption, evidence is emerging which suggests more specific targets may exist. Such specific targets may vary between organisms, thus explaining the more narrow range of activity of some essential oils and/or components. In such cases, specificity may be related to individual essential oil components. Recently, investigators attempted to determine the mechanism of action of cold-pressed Valencia orange oil against methicillin-resistant *Staphylococcus aureus* (MRSA) [32]. Microarray data showed a 24-fold increase in expression of *cwrA* following exposure to the oil. Interestingly, upregulation of *cwrA* was also demonstrated following exposure to known cell wallactive antibiotics such as penicillin G, oxacillin, phosphomycin, imipenem, and vancomycin suggesting a similar mechanism of action [33-35]. Other specific effects of citrus oil on MRSA include increased expression of penicillin-binding-protein-4 (PBP 4), involved in peptidoglycan synthesis, and genes in the *dltABCD* operon. This operon controls alanylation of teichoic acids of the cell wall which may play a role in autolysin activity of *S. aureus* [32]. Autolysin activity was also suggested by Carson and coworkers who noted that tea tree oil resulted in release of membrane-bound, cell wall autolytic enzymes leading to cell lysis and death [21]. Specific targets have also been implicated by the differential activity of essential oils observed against various microorganisms [12]. For instance, multiple studies have shown that essential oils work well against a number of Gram-positive bacteria, with only moderate to little effect on Gram-negative organisms [12]. Some investigators postulated that Gram-negative organisms were intrinsically more resistant to the effects of essential oils due to the presence of the outer membrane which provides an additional permeability barrier [36]. However, susceptibility of Gram-negative bacteria can vary by genus and species. *Aeromonas*

hydrophila, a Gram-negative bacteria commonly found in water, was highly susceptible to the effects of essential oils via an unknown mechanism; *Enterobacter aerogenes* was inhibited by cinnamon oil via interaction with various amino acid decarboxylases [37-40]. In these examples, the difference in susceptibility may be due to the presence or absence of the essential oil-specific target versus other Gram-positive or -negative bacteria; alternatively, the specific target may be present but exist in a different isoform resulting in altered susceptibility. Other specific mechanisms of action have been identified which involve quorum sensing, cellular division, sporulation, stress responses and efflux pumps. Many Gram-positive and -negative bacterial organisms communicate in a complex interplay known as 'quorum sensing' which is used to regulate various cellular functions ranging from biofilm formation and swarming to expression of virulence factors and toxins [12]. It has been suggested that interruption of these bacterial communication networks may inhibit attachment and invasion by some pathogens exploiting an alternative pathway for antimicrobial development as compared with current antibiotics [41, 42]. Interference of quorum sensing has been demonstrated by a number of plant extracts, including garlic, which resulted in significant inhibition of biofilm formation in *P. aeruginosa* [43,44]. This inhibition not only appeared to be concentration dependent, but also illustrated properties of competitive binding as suggested by structure-activity relationship studies [43,44]. Biofilm formation was also inhibited in *S. aureus* and *Salmonella enterica* serovar *typhimurium* following exposure to carvacrol, a monoterpene found in many essential oils [45]. These findings suggest inhibition of quorum sensing and biofilm formation may provide unique and as yet, unexplored targets for development of new antibiotics. However, other new drug targets may exist, which disrupt cellular division and sporulation as observed with filamentous fungi exposed to various essential oils [46]. In 2006, Pawar and Thaker [45] demonstrated that cinnamon bark oil was highly active against *Aspergillus niger* resulting in reduced production of hyphae and spores and in some cases complete inhibition of growth. The underlying mechanism(s) for these observations were not determined. However, previous investigators identified a correlation between inhibition of sporulation and cellular respiration versus growth [47]. Specifically, essential oils such as citron and lavender significantly inhibited sporulation and cellular respiration, with little effect on growth, whereas oils from cinnamon bark and lemongrass decreased growth, with little to no effect on sporulation or cellular respiration [47]. The effect on cellular respiration has implications for additional drug targets, especially those involving energy-dependent processes such as efflux of various macromolecules as seen with bacterial efflux pumps. Bacterial efflux pumps are responsible for multidrug resistance in a number of bacteria including the

AcrAB-TolC efflux system in the *Enterobacteriaceae* and the MexAB-OprM system in *Pseudomonas aeruginosa* [12]. Recent evidence suggests that these efflux mechanisms may in part be responsible for the decreased susceptibility of many Gram-negative organisms to plant-derived phytochemicals and essential oils. However, some oils such as falcariindiol, derived from *Levisticum officinale*, and the geraniol containing *Helicrysum italicum* have demonstrated anti-efflux activity especially in combination with ciprofloxacin and chloramphenicol, respectively, against Gram-negative bacteria [48,49].

Other common components of essential oils with specific antimicrobial activity are alcohols and aldehydes. Alcohols, especially the terpene alcohols, have significant bactericidal activity against a wide range of microorganisms. This bactericidal activity is thought to occur via a number of mechanisms including denaturation of proteins, dehydration of bacterial cells, or solvation of bacterial cell membranes [50,51]. In comparison, aldehydes are thought to interfere with reactions involving electron transfer, especially when conjugated to a carbon-carbon double bond. Such an electronegative molecular arrangement would result in interference with a large number of biological reactions of central metabolism (e.g. respiration and carbon cycling) resulting in rapid cell death [51].

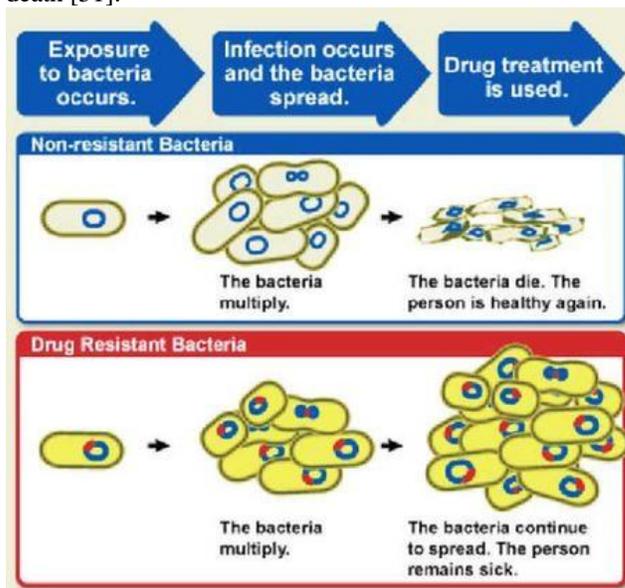


Fig 3.

Natural Remedies and antibiotic resistant superbugs

Research and development of new antibiotics decreased significantly in the 1970's when the need for new drugs was thought to be negligible since infectious diseases were becoming a concern of the past. As a result, when new antibiotics were needed (e.g. when resistance emerged), pharmaceutical companies merely modified existing antibiotics via slight structural alterations. This

approach was more economical than developing a completely new drug, especially at a time when the prevailing perception was that humanity had conquered infectious disease [52]. Today, infections have been documented which are resistant to all known drugs; treatment is often problematic and unsuccessful [53]. Unfortunately, antibiotics of ‘last resort’ are often used, including drugs previously abandoned due to overt toxicity or serious side effects [54]. Yet even this approach fails to offer long-term solutions for emerging microbial resistance to existing agents and prevention of resistance to new drugs. Perhaps what is needed is a paradigm shift, a fundamental alteration of the way we use antibiotics to treat infectious diseases. In this regard, there are lessons to be learned from plants. For example, plants produce a number of antimicrobial compounds including a large number of essential oils. These essential oils are comprised of numerous compounds which vary in potency and spectrum of activity both individually and as mixtures. Plants need this diversity considering the variability in microbial threats encountered in the environment. Thus, essential oils often inhibit a wide range of microbes due to the synergy afforded by individual components against multiple bacterial targets. Likewise, synergy has been documented between existing antibiotics with specific combinations utilized heavily in current medical practice (e.g. trimethoprim/sulfamethoxazole; amoxicillin/clavulanate; piperacillin/ tazobactam) [55]. However, synergy between existing antibiotics and essential oils and/or components has not been thoroughly investigated; although to date, limited studies have been conducted [56]. For example, β -lactam antibiotics inhibit cell wall synthesis through interaction with penicillin-binding proteins (PBP’s) [57]. PBP2a, is a specific PBP in *S. aureus* with reduced affinity for β -lactam antibiotics resulting in resistance to these drugs [58]. Interestingly, when β -lactam antibiotics were combined *in vitro* with corilagin, a polyphenol derived from *Arctostaphylos uva-ursi*, the PBP2a-mediated resistance in MRSA was overcome with a concomitant reduction in MIC [59]. It is postulated that corilagin may interfere with binding of β -lactams to the PBP2a enzyme resulting in reversion of resistance [60]. Other plant derived compounds from green tea demonstrated a similar effect in a dose-dependent manner suggesting the presence of a specific target [61]. Synergy has also been documented with linalool and α -terpineol from *Melaleuca leucodendron* when combined with ampicillin and kanamycin [62]. In addition, synergy was seen with totarol, ferulenol, and plumbagin in combination with isoniazid (INH) and rifampin (RIF) against *Mycobacterium tuberculosis* (MTB). These combinations increased the potency of INH 4-fold against MTB [62]. Another compound isolated from the roots of

Euclea natalensis decreased the MIC 4- to 6-fold for INH and RIF, respectively [63]. Taken together, these are important findings due to the rapid emergence of multidrug-resistant tuberculosis (MDR-TB) and extensively drug-resistant tuberculosis (XDR-TB). MDR-TB is defined as resistance to INH and RIF; XDR-TB is defined as resistance to INH, RIF, and any of the fluoroquinolones and one of the injectable second-line drugs (e.g. capreomycin, amikacin, or kanamycin) [64,65]. Unfortunately, these drug-resistant patterns in MTB may become “obsolete” in the near future, as MTB strains with alarming and more extensive resistance patterns have been isolated from multiple locations on the globe. These strains exhibited resistance to nearly all drugs ever used for treatment of tuberculosis and other mycobacterial infections including: INH, RIF, ethambutol, pyrazinamide, ofloxacin, moxifloxacin, capreomycin, kanamycin, amikacin, para-aminosalicylic acid, ethionamide, cycloserine, rifabutin, clofazimine, dapsone, clarithromycin, and thiacetazone [64]. Although consensus is lacking for a specific acronym for describing these strains (extremely- versus totally-drug-resistant TB; XXDR and TDR, respectively), the fact that they have been isolated is cause for great concern. In the absence of new antibiotics becoming quickly available for treatment, an alternative approach may be to combine existing drugs with essential oils. Yet, viable combinations will require a significant investment to better understand the mechanism of action of essential oils and components, determine individual and combined toxicity, characterize metabolism *in vivo*, as well as define their selectivity and bioavailability.

SUMMARY AND CONCLUSION

Essential oils and their constituents have been used to treat a large number of human illnesses. Today, essential oils are used in alternative and holistic medicine for similar purposes and administered orally, topically or via aromatherapy. A growing number of scientific investigators have begun the process of elucidating the specific mechanism(s) of action of essential oils and components.

Emerging evidence has shown that many essential oils have both nonspecific and specific mechanisms of action which varies based on the relative abundance and chemical composition of the components.

Elucidation of the mechanism of action of these compounds may enable identification of new antibiotic targets and exploitation of novel biochemical pathways; pathways not currently targeted by existing antibiotics. combination of existing drugs with essential oils and/or components may provide an alternative approach to combat emerging drug resistance.

REFERENCES

1. Cockburn TA. The evolution and eradication of infectious diseases. *Perspect Biol Med*, 7, 1964, 498-499.

2. Anonymous. <http://www.cdc.gov/mmwr/preview/mmwrhtml/mm6209>.
3. Anonymous. <http://tngmai.files.wordpress.com/2012/09/handbookofessentionaloil.pdf>.
4. Gurib-Fakim A. Medicinal Plants: Traditions of Yesterday and Drugs of Tomorrow. *Mol Aspects Med*, 27,2006, 1-93.
5. Vollesen K, Hedberg I, Edwards S Flora of Ethiopia. Addis Ababa, Ethiopia. *National Herbarium*, 3,1989,442-478.
6. Anonymous. <http://www.itmonline.org/arts/myrrh.htm>
7. Tucker AO. Frankincense and Myrrh. *Econ Bot*, 40, 1986, 425-433.
8. Yen KY. *The Illustrated Chinese Materia Medica* Crude and Prepared. 1992, SMC Publishing Inc. Taipei.
9. Nomicos EY. Myrrh: medical marvel or myth of the Magi? *Holist Nurs Pract*, 21, 2007, 308-323.
10. Anonymous. http://tcm.chinese.cn/en/article/2009-08/24/content_10979.htm
11. Worwood VA .*The Complete Book of Essential Oils and Aromatherapy: Over 600 Natural, Non-Toxic and Fragrant Recipes to Create Health - Beauty - a Safe Home Environment* (1stedn), New World Library 1991.
12. Burt S. Essential oils: their antibacterial properties and potential applications in foods--a review. *Int J Food Microbiol*, 94, 1991, 223-253.
13. Bakkali F, Averbeck S, Averbeck D, Idaomar M. Biological effects of essential oils--a review. *Food Chem Toxicol*, 46, 2008, 446-475.
14. Baser KHC, Demirci F .Essential Oils. In Kirk-Othmer *Encyclopedia of Chemical Technology*, 4th Edition, Wiley, 2011, 1-37.
15. Baser KHC, Demirci F. Chemistry of essential oils. In *flavours and fragrances: chemistry, bioprocessing and sustainability*. Berger RG (ed) Springer, Berlin, 2007, 43-86.
16. Viljoen A, van Vuuren S, Ernst E, Klepser M, Demirci B, et al. Osmitopsis asteriscoides (Asteraceae)-the antimicrobial activity and essential oil composition of a Cape-Dutch remedy. *J Ethnopharmacol*, 88, 2003, 137-143.
17. Baser KH. Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils. *Curr Pharm Des*, 14, 2008, 3106-3119.
18. Cox SD, Mann CM, Markham JL, Bell HC, Gustafson JE, et al. The mode of antimicrobial action of the essential oil of *Melaleuca alternifolia* (tea tree oil). *J Appl Microbiol*, 88, 2000, 170-175.
19. Fisher K, Phillips CA. The effect of lemon, orange and bergamot essential oils and their components on the survival of *Campylobacter jejuni*, *Escherichia coli* O157, *Listeria monocytogenes*, *Bacillus cereus* and *Staphylococcus aureus* in vitro and in food systems. *J Appl Microbiol*, 101, 2006, 1232-1240.
20. Skandamis PN, Nychas GJ.Effect of oregano essential oil on microbiological and physico-chemical attributes of minced meat stored in air and modified atmospheres. *J Appl Microbiol*, 91, 2991, 1011-1022.
21. Carson CF, Mee BJ, Riley TV. Mechanism of action of *Melaleuca alternifolia* (tea tree) oil on *Staphylococcus aureus* determined by time-kill, lysis, leakage, and salt tolerance assays and electron microscopy. *Antimicrob Agents Chemother*, 46, 2002, 1914-1920.
22. Knobloch K, Weigand H, Weis N, Schwarm HM, Vogenschow H. Action of terpenoids on energy metabolism. In Brunke EJ (ed) *Progress in essential oil research*. Berlin, Germany, Walter de Gruyter and Co, 1986, 429-445.
23. Sikkema J, de Bont JA, Poolman B. Interactions of cyclic hydrocarbons with biological membranes. *J Biol Chem*, 269, 1994, 8022-8028.
24. Prabuseenivasan S, Jayakumar M, Ignacimuthu S. In vitro antibacterial activity of some plant essential oils. *BMC Complement Altern Med*, 6, 2006, 39.
25. Fisher K, Phillips C. The mechanism of action of a citrus oil blend against *Enterococcus faecium* and *Enterococcus faecalis*. *J Appl Microbiol*, 106, 2009, 1343-1349.
26. Oosterhaven K, Poolman B, Smid EJ. S-carvone as a natural potato sprout inhibiting, fungistatic and bacteristatic compound. *Industrial Crops and Products*, 4, 1995, 23-31.
27. Denyer SP, Hugo WB. Mechanisms of antibacterial action: a summary. In: Denyer SP, Hugo WB (Eds) *Mechanisms of Action of Chemical Biocides*. Blackwell, Oxford, 1991, 331-334.
28. Sikkema J, de Bont JA, Poolman B. Mechanisms of membrane toxicity of hydrocarbons. *Microbiol Rev* 59,1995, 201-222.
29. Davidson PM. Chemical preservatives and natural antimicrobial compounds. In: Doyle, MP, Beuchat LR, Montville TJ (Eds), *Food Microbiology, Fundamentals and Frontiers*, ASM, Washington, 1997, pp. 520-556.
30. Juven BJ, Kanner J, Schved F, Weisslowicz H. Factors that interact with the antibacterial action of thyme essential oil and its active constituents. *J Appl Bacteriol*, 76, 1994, 626-631.
31. Ultee A, Smid EJ. Influence of carvacrol on growth and toxin production by *Bacillus cereus*. *Int J Food Microbiol*, 64, 2001, 373-378.
32. Muthaiyan A, Martin EM, Natesan S, Crandall PG, Wilkinson BJ, et al. Antimicrobial effect and mode of action of terpenless cold-pressed Valencia orange essential oil on methicillin-resistant *Staphylococcus aureus*. *J Appl Microbiol*, 112, 2012, 1020-1033.

33. McAleese F, Wu SW, Sieradzki K, Dunman P, Murphy E, et al. Overexpression of genes of the cell wall stimulon in clinical isolates of *Staphylococcus aureus* exhibiting vancomycin-intermediate- *S. aureus*-type resistance to vancomycin. *J Bacteriol*, 188, 2006, 1120-1133.
34. Sobral RG, Jones AE, Des Etages SG, Dougherty TJ, Peitzsch RM, et al. Extensive and genome-wide changes in the transcription profile of *Staphylococcus aureus* induced by modulating the transcription of the cell wall synthesis gene *murF*. *J Bacteriol*, 189, 2007, 2376-2391.
35. Balibar CJ, Shen X, McGuire D, Yu D, McKenney D, et al. *cwr A*, a gene that specifically responds to cell wall damage in *Staphylococcus aureus*. *Microbiology*, 156, 2010, 1372-1383.
36. Vaara M. Agents that increase the permeability of the outer membrane. *Microbiol Rev* 56, 1992, 395-411.
37. Deans SG, Ritchie G. Antibacterial properties of plant essential oils. *Internatl J Food Microbiol* 5, 1987, 165-180.
38. Hao YY, Brackett RE, Doyle MP. Inhibition of *Listeria monocytogenes* and *Aeromonas hydrophila* by plant extracts in refrigerated cooked beef. *J Food Prot* 61, 1998, 307-312.
39. Wendakoon CN, Sakaguchi M. Inhibition of amino acid decarboxylase activity of *Enterobacter aerogenes* by active components in spices. *J Food Protection* ,58, 1995, 280-283.
40. Williams P. Quorum sensing, communication and cross-kingdom signalling in the bacterial world. *Microbiology*, 153,2007, 3923-3938.
41. Zucca M, Crivellaro S, Savoia D. New trends in the inhibition of *Pseudomonas aeruginosa* quorum sensing activity. In *Cystic Fibrosis: Etiology, Diagnosis, and Treatments. Leatte PN*, Nova Publishing, NY.2009.
42. Rasmussen TB, Skindersoe ME, Bjarnsholt T, Phipps RK, Christensen KB, et al. Identity and effects of quorum-sensing inhibitors produced by *Penicillium* species. *Microbiology*, 151, 2005, 1325-1340.
43. Rasmussen TB, Bjarnsholt T, Skindersoe ME, Hentzer M, Kristoffersen P, et al. Screening for quorum-sensing inhibitors (QSI) by use of a novel genetic system, the QSI selector. *J Bacteriol* ,187, 2005, 1799-1814.
44. Knowles JR, Roller S, Murray DB, Naidu AS. Antimicrobial action of carvacrol at different stages of dual-species biofilm development by *Staphylococcus aureus* and *Salmonella enterica* serovar Typhimurium. *Appl Environ Microbiol*, 71, 2005, 797-803.
45. Pawar VC, Thaker VS . In vitro efficacy of 75 essential oils against *Aspergillus niger*. *Mycoses*, 49, 2006, 316-323.
46. Inouye S, Watanabe M, Nishiyama Y, Takeo K, Akao M, et al. Antisporulating and respiration-inhibitory effects of essential oils on filamentous fungi. *Mycoses*, 41, 1998, 403-410.
47. Lorenzi V, Muselli A, Bernardini AF, Berti L, Pagès JM, et al. Geraniol restores antibiotic activities against multidrug-resistant isolates from gramnegative species. *Antimicrob Agents Chemother*, 53, 2009, 2209-2211.
48. Garvey MI, Rahman MM, Gibbons S, Piddock LJ. Medicinal plant extracts with efflux inhibitory activity against Gram-negative bacteria. *Int J Antimicrob Agents*, 37, 2011, 145-151.
49. Pelczar MJ, Chan ECS, Krieg NR. Control of microorganisms, the control of microorganisms by physical agents. *In Microbiology*, New York, McGraw-Hill International, 1998
50. Dorman HJ, Deans SG. Antimicrobial agents from plants: antibacterial activity of plant volatile oils. *J Appl Microbiol* ,88,2000,308-316.
51. Langeveld WT, Veldhuizen EJ, Burt SA. Synergy between essential oil components and antibiotics: a review. *Crit Rev Microbiol*, 2013
52. Udawadia ZF, Amale RA, Ajbani KK, Rodrigues C. Totally drug-resistant tuberculosis in India. *Clin Infect Dis*, 54, 2012, 579-581.
53. Boucher HW, Talbot GH, Bradley JS, Edwards JE, Gilbert D, et al. Bad bugs, no drugs: no ESCAPE! An update from the Infectious Diseases Society of America. *Clin Infect Dis*, 48, 2009, 1-12.
54. Bush K, Courvalin P, Dantas G, Davies J, Eisenstein B, et al. Tackling antibiotic resistance. *Nat Rev Microbiol*, 9, 2011, 894-896.
55. Hemaiswarya S, Kruthiventi AK, Doble M. Synergism between natural products and antibiotics against infectious diseases. *Phytomedicine*, 15, 2008, 639- 652.
56. Ghuysen JM. Molecular structures of penicillin-binding proteins and beta-lactamases. *Trends Microbiol*, 2, 1994, 372-380.
57. Lu WP, Sun Y, Bauer MD, Paule S, Koenigs PM, et al. Penicillinbinding protein 2a from methicillin-resistant *Staphylococcus aureus*: kinetic characterization of its interactions with beta-lactams using electrospray mass spectrometry. *Biochemistry*, 38, 1999, 6537-6546.
58. Shimizu M, Shiota S, Mizushima T, Ito H, Hatano T, et al. Marked potentiation of activity of beta-lactams against methicillin-resistant *Staphylococcus aureus* by corilagin. *Antimicrob Agents Chemother*, 45, 2001, 3198- 3201.
59. Shiota S, Shimizu M, Sugiyama J, Morita Y, Mizushima T, et al. Mechanisms of action of corilagin and tellimagrandin I that remarkably potentiate the activity of beta-lactams against methicillin-resistant *Staphylococcus aureus*. *Journal of Microbiol Immunol*, 48, 2004, 67-73.

60. Zhao W, Hu Z, Hara Y, Shimamura T. Inhibition of penicillinase by epigallocatechin-gallate resulting in restoration of antibacterial activity of penicillin against penicillinase-producing *Staphylococcus aureus*. *Antimicrob Agents Chemother* 46, 2002, 2266-2268.
61. Avenirova EL, Ashmarin IP, Movchan NA, Lapina IK. Combination of novoimanine with antibiotics with a different mechanism of action. *Antibiotiki*, 20, 1975, 636-639.
62. Mossa JS, El-Ferally FS, Muhammad I. Antimycobacterial constituents from *Juniperus procera*, *Ferula communis* and *Plumbago zeylanica* and their in vitro synergistic activity with isonicotinic acid hydrazide. *Phytother Res*, 18, 2004, 934-937.
63. Bapela NB, Lall N, Fourie PB, Franzblau SG, Van Rensburg CE. Activity of 7-methyljuglone in combination with antituberculous drugs against *Mycobacterium tuberculosis*. *Phytomedicine*, 13, 2006, 630-635.
64. Migliori GB, De Iaco G, Besozzi G, Centis R, Cirillo DM. First tuberculosis cases in Italy resistant to all tested drugs. *Euro Surveil*. 12, 2007, E070517.
65. Extensively drug-resistant tuberculosis (XDR-TB): recommendations for prevention and control. *Wkly Epidemiol Rec*, 81, 2006, 430-432.